

# Lactate and Risk of Incident Diabetes in a Case-Cohort of the Atherosclerosis Risk in Communities (ARIC) Study

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## Abstract

**Background:** Oxidative capacity is decreased in type 2 diabetes. Whether decreased oxidative capacity is a cause or consequence of diabetes is unknown. Our purpose is to evaluate whether lactate, a marker of oxidative capacity, is associated with incident diabetes.

**Methods and Findings:** We conducted a case-cohort study in the Atherosclerosis Risk in Communities (ARIC) study at year 9 of follow-up. We evaluated lactate's association with diabetes risk factors at baseline and estimated the hazard ratio for incident diabetes by quartiles of plasma lactate in 544 incident diabetic cases and 533 non-cases. Plasma lactate showed a graded positive relationship with fasting glucose and insulin ( $P < 0.001$ ). The relative hazard for incident diabetes increased across lactate quartiles ( $P$ -trend  $\leq 0.001$ ). Following adjustment for demographic factors, medical history, physical activity, adiposity, and serum lipids, the hazard ratio in the highest quartile was 2.05 times the hazard in the lowest quartile (95% CI: 1.28, 3.28). After including fasting glucose and insulin the association became non-significant.

**Conclusions:** Lactate, an indicator of oxidative capacity, predicts incident diabetes independent of many other risk factors and is strongly related to markers of insulin resistance. Future studies should evaluate the temporal relationship between elevated lactate and impaired fasting glucose and insulin resistance.

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## Introduction

In the United States, the prevalence of type 2 diabetes is high [1] and its incidence is increasing due to rising levels of adiposity. Prior to the clinical diagnosis of diabetes, individuals undergo a period of subclinical insulin resistance [2]. Accumulating evidence suggests that low oxidative capacity due to impaired mitochondrial oxidative phosphorylation [3] is associated with insulin resistance and type 2 diabetes. It is unclear, however, whether low oxidative capacity is a cause or consequence of diabetes [4]. This question can be addressed using markers of oxidative capacity in longitudinal studies of incident diabetes.

Blood lactate is a measure of the gap between energy expenditure and oxidative capacity. Lactate is produced by

anaerobic glycolysis in muscle, adipose, and other tissues. Anaerobic glycolysis, and, therefore, lactate production increase when energy demand exceeds mitochondrial oxidative capacity. Traditionally, blood lactate is used as a clinical marker of ischemia and as a measure of fitness among exercising individuals. Lactate variation in resting individuals may also be informative, however. Elevated lactate may be observed with mitochondrial impairments in oxidative phosphorylation [5,6] and several clinical studies have demonstrated that lactate is closely related to insulin resistance [7–9]. In addition, a number of observational studies have shown that lactate is associated fasting glucose, fasting insulin, and the prevalence of type 2 diabetes [10–12]. These studies, however, were limited by cross-sectional designs. Only one prospective study in white men studied the association between plasma lactate levels

and incident diabetes. This study showed that the highest quintile of lactate at baseline was associated with 2.4 (95% CI: 1.0–5.9) times the risk of developing diabetes in the lowest quintile [13]. Therefore, we examined the longitudinal association of plasma lactate with subsequent development of type 2 diabetes in a biracial cohort of middle-aged men and women using a case-cohort design.

## Materials and Methods

### Study Design

ARIC is a community-based prospective cohort study of 15,792 adults, ages 45–64. ARIC participants were identified by probability sampling from 4 U.S. communities (Forsyth County, North Carolina; Jackson, Mississippi; suburban Minneapolis, Minnesota; and Washington County, Maryland) and were enrolled between 1987 and 1989 [14–16]. Study participants returned for three follow-up visits in 1990–1992, 1993–1995, and 1996–1998, during which time incident diabetes was determined. Response rates for each ARIC visit were 60% (visit 1), 93% (visit 2), 86% (visit 3), and 80% (visit 4). The study protocol was approved by institutional review boards at each clinical site, namely, the IRBs of the University of Minnesota, Johns Hopkins University, University of North Carolina, University of Mississippi Medical Center, and Wake Forest University. Written consent was obtained from all ARIC participants.

We used a case-cohort design described previously [17]. We excluded participants with a diagnosis of diabetes at baseline ( $n = 2,018$ ), members of ethnic groups with a small sample size ( $n = 95$ ), participants whose diabetes status could not be determined in any of the three follow-up visits ( $n = 879$ ), participants lacking a valid diabetes determination ( $n = 26$ ), participants with missing plasma samples, anthropometric data, or related covariate data (hypertension, smoking, etc.) at baseline ( $n = 2,526$ ), and non-fasting individuals at baseline ( $n = 212$ ). Of the remaining 9,740 subjects, 1,105 developed diabetes during the 9-yr follow-up period. From this population, we selected a random cohort sample ( $n = 637$ ) and a random incident diabetes case sample ( $n = 552$ ) according to strata determined by race. Among those in the random cohort sample, 94 (14.8%) developed diabetes during the follow-up period. Of this population, an additional 7 non-case participants in the random cohort sample and 11 participants in the diabetes case sample did not have a lactate measurement and were excluded. After these exclusions, baseline covariates were compared between both the case-cohort and the complete cohort of ARIC visit 1 and found to be nearly identical.

### Plasma Lactate

We measured plasma lactate in stored samples originally collected in 1987–1989 and frozen at  $-70^{\circ}\text{C}$ . Lactate was quantified with a Roche Hitachi 911 auto-analyzer, using an enzymatic reaction that converts lactate to pyruvate [18]. Quality control assessments of the assays suggested an inter-assay coefficient of reliability of 0.93 and a coefficient of variation of 9.2% [12].

### Primary Outcome: Incident Diabetes

Participants were assessed for a diagnosis of diabetes at baseline and at each of the three follow-up visits. Incident diabetes was classified as a diabetic case if a participant fulfilled any one of the following conditions: (1) a fasting glucose  $\geq 7.0$  mmol/liter, (2) a nonfasting glucose  $\geq 11.1$  mmol/liter, (3) use of a diabetes medication, or (4) self-reported physician diagnosis. For individuals diagnosed by a glucose measurement, the date of diagnosis

was estimated under the assumption that glucose increased at a linear rate between visits. This linear approximation was used to estimate the time of actual diabetes onset. Similarly, the date of diagnosis for cases identified by medication use or physician diagnosis was estimated assuming a linear increase in glucose from their last diabetes-free visit [19]. As a sensitivity analysis, we restricted our diagnosis of incident diabetes to diabetes medication use or self-report of physician diagnosis, excluding the subclinical cases of diabetes identified by glucose measures.

### Other Covariates

We included a number of covariates assessed at the baseline visit of ARIC that were known to be associated with diabetes. Definitions and methods for the assessment of these covariates were described previously [20,21]. In brief, the following laboratory measurements were treated as continuous variables: low density lipoprotein (LDL) cholesterol, high density lipoprotein (HDL) cholesterol, triglycerides, fasting glucose, and fasting insulin. Triglycerides and insulin were log-base 10 transformed to normalize their distributions. We also determined the homeostatic model assessment insulin resistance index (HOMA-IR) by  $((\text{fasting insulin in pmol/L})/6.945 * (\text{fasting glucose in mg/dL}))/405$ .

With regard to physical examination covariates, we determined body mass index ( $\text{kg/m}^2$ ) from height and weight measurements and measured waist circumference at the umbilical level. Smoking status (never, current, former), education ( $<12$  years,  $\geq 12$  years), and leisure time physical activity index (ordinal integer ranging from 1 to 4) were based on replies to the Baecke Physical Activity questionnaire. Prevalent coronary heart disease (yes or no) was based on past history of myocardial infarction, heart or arterial surgery, coronary bypass, or angioplasty. Parental history of diabetes (yes or no) was defined by a diagnosis of diabetes in a participant's father or mother. Hypertension (yes or no) was defined as either a systolic blood pressure measurement  $\geq 140$  mmHg, a diastolic blood pressure measurement  $\geq 90$  mmHg, or use of high blood pressure medications in the past 2 weeks.

### Statistical Analysis

All analyses and plots were weighted to account for the random cohort sample and the random case sample using the `svy` command and `pweights` in Stata 11.1 (StataCorp LP, College Station, TX), which entail inverse weighting of the observations according to the case-cohort sampling design [20]. Proportions and mean baseline characteristics of the study participants were reported by lactate quartiles, where lactate quartile categories were based on measurements in the random cohort sample. Standard errors were estimated using the Taylor series (linearization) method. We performed a cross-sectional analysis of the association between log-base 10 transformed lactate and physiologic correlates of insulin resistance, namely, BMI, waist circumference, log-base 10 transformed triglycerides, HDL cholesterol, fasting glucose, and log-base 10 transformed fasting insulin. Comparisons were made using linear regression and Pearson's correlation coefficients. The HOMA-IR was plotted by lactate quartiles to further examine the relationship between lactate and lipids or glycemia. The  $P$ -value for the trend between HOMA-IR and lactate quartiles were calculated with linear regression, treating the median value of each lactate quartile as an ordinal variable.

A Kaplan-Meier cumulative incidence plot was utilized to visualize the relationship between baseline lactate measurements and risk of incident diabetes, using follow-up time as the time axis. A Wald test was used to evaluate for a trend across quartiles of

lactate. The association between lactate quartiles and incident diabetes was further examined in nested Cox proportional hazards models that (1) addressed factors associated with both diabetes and lactate, and (2) explored the metabolic pathway relating lactate with incident diabetes by adjusting for the same factors above along with insulin and glucose. In the first series of models, we adjusted for age, gender, race, ARIC center, and education (Model 1); a diagnosis of hypertension, prevalent coronary heart disease, smoking status, leisure index, and parental history of diabetes (Model 2); BMI and waist circumference (Model 3); and log10-transformed triglycerides, LDL cholesterol, and HDL cholesterol (Model 4). In our metabolic pathway models, we adjusted for fasting glucose (Model 5a), fasting insulin (Model 5b) and both (Model 5c) in addition to the preceding covariates. Furthermore, we generated histograms of the distribution of lactate among diabetic cases and noncases and overlaid a restricted cubic spline based on Model 4 with knots at each lactate quartile to visualize the risk association.

## Results

The geometric mean for plasma lactate in the entire study sample ( $N = 1,077$ ) was 7.67 mg/dL (IQR: 5.80 to 9.70 mg/dL; max: 36.1 mg/dL) (**Table 1**); plasma lactate was less than 18 mg/dL ( $\sim 2$  mmol/L) in 97% of the population. Age was not related to lactate concentration ( $P = 0.55$ ). However, the proportion of African Americans and men increased across lactate quartiles ( $P$ -trends of 0.03 and  $<0.001$ , respectively). The prevalence of hypertension was also greater in higher lactate quartiles, ranging from 13.9% to 34.2% ( $P$ -trend  $<0.001$ ). Lifestyle characteristics, i.e. leisure physical activity index, smoking status, or education attainment, prevalent coronary heart disease, and parental history of diabetes were not associated with lactate quartiles.

Higher lactate was associated with both BMI and waist circumference (both  $P < 0.001$ ) (**Table 1**). With regard to lipids, LDL cholesterol did not vary significantly across lactate quartiles ( $P = 0.38$ ), while triglycerides concentrations were greater, and HDL levels lower, with successive lactate quartiles ( $P$  for trend of  $<0.001$  and 0.04, respectively) (**Table 1**). Fasting glucose and fasting insulin also demonstrated significant positive trends by lactate quartile (both  $P < 0.001$ ) (**Table 1**). In linear regression, BMI, waist circumference, triglycerides, fasting glucose and fasting insulin were also positively associated with plasma lactate, while HDL cholesterol was negatively associated with plasma lactate (**Table 2**). Similarly, the HOMA-IR (**Figure 1**) was higher with successive lactate quartiles ( $P < 0.001$ ).

There was a graded relationship between plasma lactate at baseline and the cumulative risk of incident type 2 diabetes over an average 7.9 years of follow-up (**Figure 2**). This graded relationship was robust after simultaneous adjustment for diabetes risk factors (**Table 3**, Models 1–4). In models adjusting for medical history, anthropometric variables and lipids, the association was attenuated but remained significant and strong (4<sup>th</sup> quartile HR 2.05; 95% CI: 1.28, 3.28;  $P$ -trend  $\leq 0.001$ ). Histogram comparison revealed an upward shift in the distribution of lactate in diabetic cases versus non-diabetics (**Figure 3**). Similarly, a restricted cubic spline demonstrated an increase in the relative hazard for diabetes with increasing lactate levels, particularly below the 75<sup>th</sup> percentile of the lactate distribution, i.e. a plasma lactate of 9.7 mg/dL.

Models examining the metabolic pathway between lactate and incident diabetes are shown in **Table 3**. The inclusion of fasting insulin as a covariate (Model 5b) attenuated the dose-response trend (4<sup>th</sup> quartile HR 1.46,  $P$ -trend = 0.06), while the inclusion of fasting glucose (Models 5a & 5c), resulted in a non-significant

reversal of the association (Model 5a: 4<sup>th</sup> quartile HR 0.77,  $P$ -trend = 0.45; Model 5c: 4<sup>th</sup> quartile HR 0.66,  $P$ -trend = 0.26).

A sensitivity analysis defining diabetes on the basis of diabetes medication use and self-report of physician diagnosis, i.e. excluding the subclinical cases identified by glucose measures, increased the magnitude of the association between lactate and incident diabetes (**Table S1**). In fact, even after adjusting for fasting insulin, lactate still demonstrated a significant trend across quartiles ( $P$ -trend = 0.02).

## Discussion

This analysis represents the largest population-based examination of the prospective relationship between lactate and type 2 diabetes. We observed a strong, graded relationship between plasma lactate and subsequent risk of incident type 2 diabetes over a 9-year follow-up period. The graded association, which was observed across the normal clinical range of lactate values, was independent of many traditional diabetes risk factors.

Prior to the current report there were several cross-sectional, clinical studies that found an association between lactate and insulin resistance [10,11] as well as diabetes [9]. Furthermore, one prospective study of 766 Swedish men, found that an elevated lactate was associated with 2.4 times the risk of diabetes [13]. However, these studies were small and not readily generalizable. Strengths of our study include its large, biethnic community-based population of men and women with high quality measures of potential confounders.

In our study, we measured blood lactate levels in a population-based sample of resting subjects after an overnight fast and found that variations in the normal range ( $<18$  mg/dl) were predictive of incident diabetes. Lactate is a product of glycolysis that rises markedly in states of hypoxia such as ischemia [22], with exercise [23], as well as with acute elevations in glucose and insulin [24,25]. In contrast, plasma lactate in the rested, fasted state represents a balance between gluconeogenesis [26] and the rate of extrahepatic glycolysis [25]. A rise in resting lactate can be observed when gluconeogenesis is disrupted by alcohol use [27] or genetic disorders [28,29]. Higher resting lactate concentrations may also be seen with increased extrahepatic glucose disposal or glycolysis [25] or with an increase in the concentration of lactate due to hypoxia [30], or oxidative insufficiency [31].

In the fasted, resting state, lactate is produced primarily by skeletal muscle and adipose tissue, and to a lesser extent by the brain, erythrocytes, liver, gut, kidney, and skin [4,25]. While skeletal muscle accounts for the majority of lactate release [24], a major determinant of variation in plasma lactate concentration is the rate of glucose disposal in extramuscular tissues [25]. This is exemplified by a comparison of obese and non-obese persons, in which greater lactate concentrations are due to increased lactate production from adipose tissue [24]. In addition to increased disposal, increased adiposity is associated with intracellular hypoxia [30] due to increased adipocyte size [32,33] and inadequate vascularization [34]. Both these factors – excess glucose disposal and increased anaerobic glycolysis contribute to elevations in fasting lactate. However, we observed a relatively low correlation between BMI or waist circumference and lactate as well as a graded relationship between lactate and incident diabetes even after adjusting for measures of adiposity. This suggests that lactate is more than a marker of BMI or waist circumference.

There is strong evidence that oxidative insufficiency, evidenced by elevated plasma lactate concentrations, is associated with diabetes risk. Studies have found smaller mitochondrial size and lower density among adults with type 2 diabetes [35,36] and

**Table 1.** Baseline (visit 1) characteristics of ARIC participants according to quartiles of plasma lactate.

	Plasma Lactate Level (mg/dL)*					P for Trend <sup>§</sup>
	≤5.8 (N = 190 <sup>†</sup> )	5.8–7.3 (N = 226 <sup>†</sup> )	7.3–9.7 (N = 329 <sup>†</sup> )	>9.7 (N = 332 <sup>†</sup> )		
Overall (N = 1,077 <sup>†</sup> )	Mean or % (SE or IQR)	Mean or % (SE or IQR)	Mean or % (SE or IQR)	Mean or % (SE or IQR)		
Plasma lactate, mg/dL <sup>‡</sup>	7.7 (5.8 to 9.7)	5.0 (4.6 to 5.5)	6.4 (6.1 to 6.9)	8.3 (7.7 to 8.8)	13.1 (10.9 to 14.7)	<0.001
Age, y	52.8 (0.2)	52.9 (0.5)	53.0 (0.5)	52.7 (0.5)	52.6 (0.5)	0.55
Black, %	19.9 (0.7)	11.8 (0.9)	19.6 (1.4)	24.0 (1.6)	23.9 (1.8)	0.03
Men, %	36.0 (2.1)	25.8 (4.0)	30.6 (4.2)	38.2 (3.9)	49.4 (4.2)	<0.001
Education beyond high school, %	43.7 (2.1)	38.2 (4.4)	49.2 (4.5)	43.5 (4.0)	44.3 (4.2)	0.67
Hypertension, %	26.3 (1.8)	13.9 (2.8)	28.5 (3.9)	28.8 (3.3)	34.2 (3.8)	<0.001
Prevalent coronary heart disease, %	2.5 (0.7)	0.0 <sup>•</sup>	2.6 (1.5)	4.1 (1.6)	3.1 (1.5)	0.10
Smoking Status						
Never, %	48.2 (2.2)	52.0 (4.6)	42.0 (4.5)	50.1 (4.0)	48.3 (4.2)	0.98
Former, %	31.4 (2.0)	27.9 (4.1)	30.56 (4.2)	32.5 (3.8)	34.4 (4.0)	0.26
Current, %	20.4 (1.7)	20.1 (3.7)	27.4 (4.0)	17.5 (2.9)	17.3 (3.1)	0.19
Leisure index	2.4 (0.02)	2.4 (0.04)	2.3 (0.05)	2.3 (0.04)	2.4 (0.04)	0.99
Parental history of diabetes, %	24.4 (1.9)	25.2 (4.0)	20.7 (3.7)	25.4 (3.6)	25.9 (3.9)	0.65
Body mass index, kg/m <sup>2</sup>	27.2 (0.2)	26.1 (0.4)	26.3 (0.4)	28.2 (0.4)	28.2 (0.4)	<0.001
Waist circumference, cm	94.8 (0.5)	92.2 (1.0)	92.4 (1.0)	96.9 (1.0)	97.4 (1.1)	<0.001
Triglycerides, mmol/L**	1.2 (0.8 to 1.7)	1.0 (0.8 to 1.3)	1.1 (0.8 to 1.4)	1.3 (0.9 to 1.7)	1.4 (0.9 to 2.0)	<0.001
LDL cholesterol level, mmol/L	3.4 (0.04)	3.4 (0.1)	3.4 (0.1)	3.6 (0.1)	3.4 (0.1)	0.38
HDL cholesterol level, mmol/L	1.4 (0.02)	1.5 (0.04)	1.4 (0.04)	1.3 (0.03)	1.4 (0.04)	0.04
Fasting glucose, mg/dL	98.2 (0.4)	94.8 (0.7)	96.8 (0.8)	99.2 (0.6)	101.8 (0.7)	<0.001
Fasting insulin, pmol/L**	57.2 (35.9 to 86.1)	42.7 (28.7 to 64.6)	47.3 (28.7 to 71.8)	70.4 (50.2 to 100.5)	73.5 (50.2 to 114.8)	<0.001

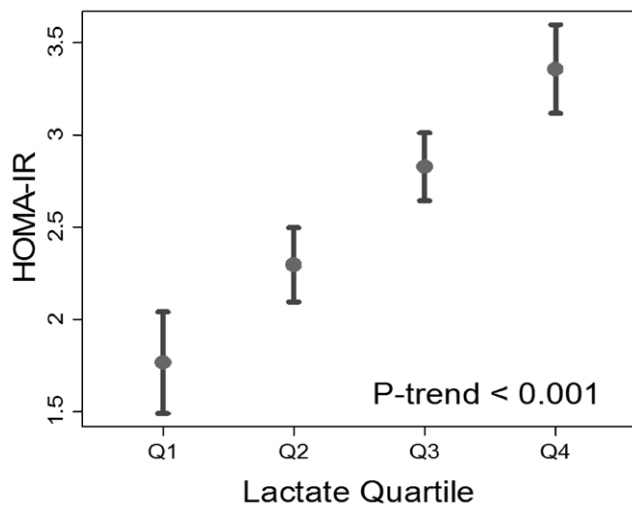
\*The ranges of the plasma lactate quartiles were determined using specimens from the weighted random cohort sample.

<sup>†</sup>Represents the maximum number of participants in each category. Actual number may vary due to missing data.<sup>‡</sup>Plasma lactate mg/dL may be converted to mmol/L by multiplying by 0.111.<sup>§</sup>p-trend evaluated with linear or logistic regression using the median lactate value for each quartile as an ordinal variable.<sup>•</sup>There were no participants with coronary heart disease in quartile 1. SE not calculated due to small sample size.

\*\*Represents geometric mean and interquartile range.

Note: LDL represents low density lipoprotein. HDL represents high density lipoprotein.

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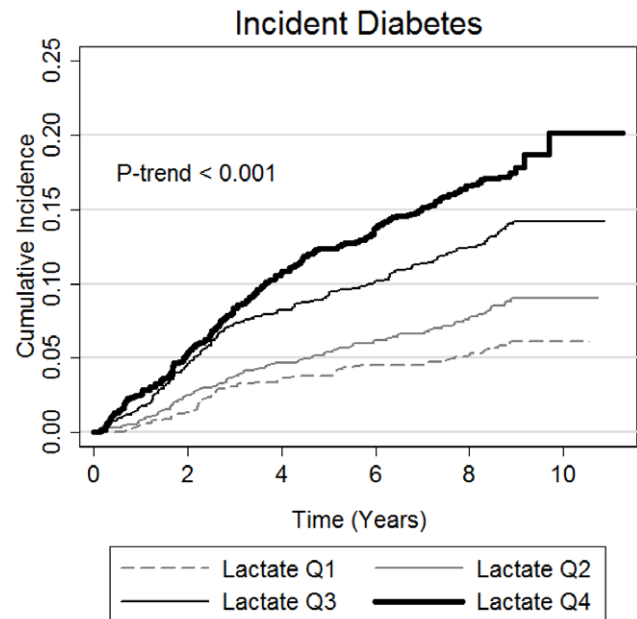


**Figure 1. Mean homeostatic model assessment (HOMA-IR) with 95% confidence intervals by baseline plasma lactate quartile.**  
doi:10.1371/journal.pone.0055113.g001

individuals with insulin resistance [37]. Furthermore, there is evidence that mitochondria are less active in diabetic adults [36,38]. Despite these associations, the temporal relationship between mitochondrial function and incident diabetes remains controversial: some argue that insulin resistance precedes and even induces mitochondrial inefficiency [39]. Since insulin signaling may improve oxidative capacity by increasing blood flow and by stimulating mitochondrial biogenesis, decreased oxidative capacity may be a consequence of decreased insulin signaling activity [40]. Decreased oxidative capacity is not universally observed in all patients with insulin resistance [41]. Furthermore, weight loss can decrease insulin resistance without impacting oxidative capacity and, among those at risk for type 2 diabetes, exercise can increase oxidative capacity without impacting insulin resistance [42–44].

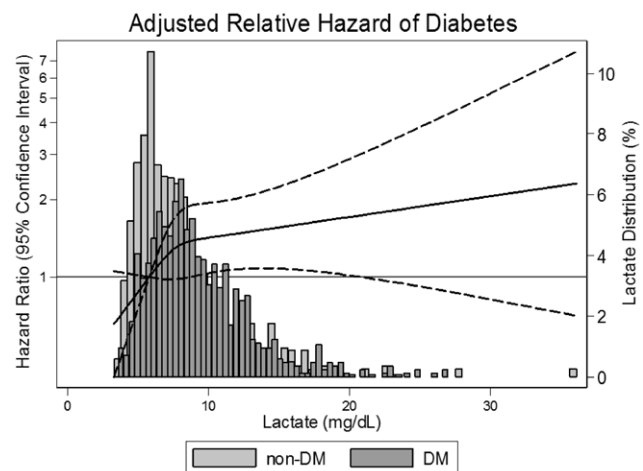
In contrast to the work just described, there is strong evidence supporting a causal role for decreased oxidative capacity in the development of insulin resistance. Experimental manipulation of the mitochondrial genome in mice results in decreased oxidative capacity and insulin resistance [45]. Similarly, genetic mutations in human mitochondrial genes are associated with insulin resistance [5,6] and diabetes mellitus [46,47]. Several studies have also shown that decreased oxidative capacity precedes insulin resistance [48]. Moreover, exercise and weight loss [49] as well as insulin sensitizers like thiazolidinediones [50] improve insulin resistance, while exerting direct effects on oxidative capacity. Our study demonstrates that lactate as a marker of decreased oxidative capacity is associated with progression of insulin resistance. However, this was not independent of concurrent baseline measures of fasting glucose and insulin, suggesting that lactate does not represent a pathway distinct from insulin resistance. While establishing the temporal relationship between lactate and insulin resistance should be an important focus of future studies, our study suggests that studying the markers in concert rather than as individual causal factors may better reflect the underlying shift in metabolism that precedes insulin resistance and ultimately diabetes.

Another potential explanation for the relationship between lactate and incident diabetes is expression and activity of GLUT4, the glucose transporter in muscle and fat cells. In response to



**Figure 2. Kaplan-Meier cumulative incidence plot with follow-up years as the time axis and incident diabetes as the outcome stratified by baseline plasma lactate value.** A Wald test was performed to assess for a trend across quartiles of lactate.  
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insulin, GLUT4 sequesters glucose into muscle and fat cells where it undergoes glycolysis [51,52]. In certain settings, greater GLUT4 expression is associated with increased lactate production [53]. Furthermore, there is evidence that lactate itself can induce insulin resistance by suppressing GLUT4 expression [54]. Confirmation of this hypothesis is beyond the scope of this study, however.



**Figure 3. Histograms comparing the distribution of lactate in diabetic cases (DM) versus noncases (non-DM).** The solid line represents a restricted cubic spline of the relative hazard with knots at 25<sup>th</sup>, 50<sup>th</sup>, and 75<sup>th</sup> percentiles. The dashed lines represent the 95% confidence interval. Model is adjusted for age, sex, race, ARIC study center, education, hypertension status, history of coronary heart disease, smoking status, leisure index, parental history of diabetes, body mass index, waist circumference, triglycerides, low density lipoprotein cholesterol, and high density lipoprotein cholesterol.  
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**Table 2.** Percent change in plasma lactate per 1 unit increase in physiologic correlates of insulin resistance and Pearson correlation coefficient between baseline characteristics and log-base 10 transformed lactate.

	Percent Change Per 1 Unit Increase*	P	Pearson's Coefficient
Body mass index, kg/m <sup>2</sup>	1.2%	<0.001	0.15
Waist circumference, cm	0.3%	<0.001	0.14
Log <sub>10</sub> (Triglycerides, mmol/L)	53.0%	<0.001	0.22
HDL cholesterol, mmol/L	−5.9%	0.04	−0.11
Fasting glucose, mg/dL	1.0%	<0.001	0.26
Log <sub>10</sub> (Fasting insulin, pmol/L)	20.3%	<0.001	0.31

\*Adjusted for age, gender, race, and center.

Note: HDL represents high density lipoprotein.

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Adjustment for insulin attenuated the association between lactate and incident diabetes, while adjustment for glucose non-significantly inverted the association. The change in direction of the association of lactate with incident diabetes after including glucose in models may be due to the role of fasting glucose in our case definition. When subclinical cases of diabetes were excluded from the outcome (**Table S1**), lactate was significantly associated with incident diabetes independently of insulin. Furthermore, lactate was no longer inversely associated with incident diabetes after adjustment for glucose, albeit non-significant. Whether this attenuation is due to loss in power from fewer cases or that lactate is not independent of glucose requires a larger study population.

**Table 3.** Hazard ratios (95% confidence intervals) for developing type 2 diabetes by weighted quartile of lactate concentrations.

	Quartile of Lactate				P for trend*
	≤5.8	5.8–7.3	7.3–9.7	>9.7	
Model 1	1.0 [Ref]	1.34 (0.89, 2.02)	2.13 (1.46, 3.10)	2.69 (1.84, 3.94)	<0.001
Model 2	1.0 [Ref]	1.19 (0.75, 1.89)	1.83 (1.21, 2.77)	2.41 (1.58, 3.68)	<0.001
Model 3	1.0 [Ref]	1.30 (0.79, 2.13)	1.45 (0.92, 2.29)	2.17 (1.38, 3.41)	<0.001
Model 4	1.0 [Ref]	1.27 (0.76, 2.13)	1.29 (0.80, 2.06)	2.05 (1.28, 3.28)	0.001
Model 5a	1.0 [Ref]	0.89 (0.51, 1.56)	0.78 (0.47, 1.31)	0.77 (0.44, 1.35)	0.45
Model 5b	1.0 [Ref]	1.05 (0.61, 1.81)	1.03 (0.64, 1.65)	1.46 (0.90, 2.39)	0.06
Model 5c	1.0 [Ref]	0.81 (0.46, 1.44)	0.70 (0.42, 1.18)	0.66 (0.37, 1.17)	0.26

Model 1: Age, gender, race, ARIC center, education.

Model 2: Model 1+ diagnosis of hypertension, prevalent coronary heart disease, smoking status, leisure index, parental history of diabetes.

Model 3: Model 2+ body mass index, waist circumference.

Model 4: Model 3+ log<sub>10</sub> triglycerides, low density lipoprotein cholesterol, high density lipoprotein cholesterol.

Model 5a: Model 4+ fasting glucose.

Model 5b: Model 4+ log<sub>10</sub> fasting insulin.Model 5c: Model 4+ fasting glucose and log<sub>10</sub> fasting insulin.

\*P-value for trend evaluated using an ordinal variable based on the median lactate in each quartile.

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Several limitations should be considered in the interpretation of these data. First, lactate was assessed with single plasma specimens that had been frozen for over 20 years. The effects of long-term storage on lactate concentration are unknown, and the lack of repeat measurements limit our ability to assess the within-person variability of lactate. Furthermore, this study is also limited with regard to the measurement of glucose and insulin. Although we attempted to explore glucose and insulin as mediators in the causal pathway of incident diabetes, our only lactate sample was measured at the same time as glucose and insulin. As a result, we were unable to establish temporality, which would be necessary to differentiate between their role as mediators or confounders of lactate and diabetes. Moreover, while we adjusted for many suspected confounders of the relationship between lactate and incident diabetes, it is possible we did not fully eliminate their effects. For example, while we adjusted for leisure index, a measure of physical activity, quantitative assessments of fitness and aerobic capacity were not performed. As a result, our models could be affected by residual confounding. Finally, it would have been informative to measure lactate during the visits between baseline and the development of diabetes as this could elucidate trends in lactate preceding and potentially contributing to incident diabetes.

In conclusion, our results show that lactate is a predictor of incident diabetes, suggesting a role for oxidative insufficiency in the pathogenesis of diabetes. Additional research is needed to determine the temporal relationship between elevated lactate and impaired fasting glucose and insulin resistance. Furthermore, future studies should attempt to elucidate the causal mechanism behind elevated lactate and diabetes pathogenesis.

## Supporting Information

**Table S1 Hazard ratios (95% confidence intervals) for developing type 2 diabetes by weighted quartile of lactate concentrations.** Type 2 diabetes defined by self-report or medication use alone (N = 233). (DOCX)

## Author Contributions

Statistical analysis: SPJ EG. Conceived and designed the experiments: SPJ GPSS AYC JHY. Performed the experiments: SPJ AYC JHY. Analyzed the data: SPJ AYC JHY. Contributed reagents/materials/analysis tools: ERM RCH CMB FLB MIS JSP. Wrote the paper: SPJ JHY.



## References

- Cowie CC, Rust KF, Byrd-Holt DD, Eberhardt MS, Flegal KM, et al. (2006) Prevalence of diabetes and impaired fasting glucose in adults in the U.S. population: National Health And Nutrition Examination Survey 1999–2002. *Diabetes Care* 29: 1263–8.
- Lillioja S, Mott DM, Spraul M, Ferraro R, Foley JE, et al. (1993) Insulin resistance and insulin secretory dysfunction as precursors of non-insulin-dependent diabetes mellitus. Prospective studies of Pima Indians. *N Engl J Med* 329: 1988–92.
- Bernal-Mizrachi C, Semenkovich CF (2006) Fast predators or fast food, the fit still survive. *Nat Med* 12: 46–47.
- Kreisberg RA (1980) Lactate homeostasis and lactic acidosis. *Ann Intern Med* 92(2 Pt 1): 227–37.
- Iossa S, Mollica MP, Lionetti L, Crescenzo R, Tasso R, et al. (2004) A possible link between skeletal muscle mitochondrial efficiency and age-induced insulin resistance. *Diabetes* 53: 2861–6.
- Petersen KF, Befroy D, Dufour S, Dziura J, Ariyan C, et al. (2003) Mitochondrial dysfunction in the elderly: possible role in insulin resistance. *Science* 300: 1140–2.
- Thorburn AW, Gumbiner B, Bulacan F, Wallace P, Henry RR (1990). Intracellular glucose oxidation and glycogen synthase activity are reduced in non-insulin-dependent (type II) diabetes independent of impaired glucose uptake. *J Clin Invest* 85: 522–9.
- Woerle HJ, Szoke E, Meyer C, Dostou JM, Wittlin SD, et al. (2006) Mechanisms for abnormal postprandial glucose metabolism in type 2 diabetes. *Am J Physiol Endocrinol Metab* 290: E67–E77.
- Chen YD, Varasteh BB, Reaven GM (1993) Plasma lactate concentration in obesity and type 2 diabetes. *Diabetes Metab* 19: 348–54.
- Lovejoy J, Newby FD, Gebhart SS, DiGirolamo M (1992) Insulin resistance in obesity is associated with elevated basal lactate levels and diminished lactate appearance following intravenous glucose and insulin. *Metab Clin Exp* 41: 22–7.
- Reaven GM, Hollenbeck C, Jeng CY, Wu MS, Chen YD (1988) Measurement of plasma glucose, free fatty acid, lactate, and insulin for 24 h in patients with NIDDM. *Diabetes* 37: 1020–4.
- Crawford SO, Hoogveen RC, Brancati FL, Astor BC, Ballantyne CM, et al. (2010) Association of blood lactate with type 2 diabetes: the Atherosclerosis Risk in Communities Carotid MRI Study. *Int J Epidemiol* 39: 1647–55.
- Ohlson LO, Larsson B, Björntorp P, Eriksson H, Svärdsudd K, et al. (1988) Risk factors for type 2 (non-insulin-dependent) diabetes mellitus. Thirteen and one-half years of follow-up of the participants in a study of Swedish men born in 1913. *Diabetologia* 31: 798–805.
- The ARIC investigators (1989) The Atherosclerosis Risk in Communities (ARIC) Study: design and objectives. *Am J Epidemiol* 129: 687–702.
- Jackson R, Chambless LE, Yang K, Byrne T, Watson R, et al. (1996) Differences between respondents and nonrespondents in a multicenter community-based study vary by gender ethnicity. *J Clin Epidemiol* 49: 1441–6.
- White AD, Folsom AR, Chambless LE, Sharret AR, Yang K, et al. (1996) Community surveillance of coronary heart disease in the Atherosclerosis Risk in Communities (ARIC) Study: methods and initial two years' experience. *J Clin Epidemiol* 49: 223–33.
- Zhu N, Pankow JS, Ballantyne CM, Couper D, Hoogveen RC, et al. (2010) High-molecular-weight adiponectin and the risk of type 2 diabetes in the ARIC study. *J Clin Endocrinol Metab* 95: 5097–104.
- Barham D, Trinder P (1972) An improved colour reagent for the determination of blood glucose by the oxidase system. *Analyst* 97: 142–5.
- Pankow JS, Duncan BB, Schmidt MI, Ballantyne CM, Couper DJ, et al. (2004) Fasting plasma free fatty acids and risk of type 2 diabetes: the atherosclerosis risk in communities study. *Diabetes Care* 27: 77–82.
- Duncan BB, Schmidt MI, Pankow JS, Ballantyne CM, Couper D, et al. (2003) Low-grade systemic inflammation and the development of type 2 diabetes: the atherosclerosis risk in communities study. *Diabetes* 52: 1799–805.
- Schmidt MI, Duncan BB, Sharrett AR, Lindberg G, Savage PJ, et al. (1999) Markers of inflammation and prediction of diabetes mellitus in adults (Atherosclerosis Risk in Communities study): a cohort study. *Lancet* 353: 1649–52.
- Arieff AI, Graf H (1987) Pathophysiology of type A hypoxic lactic acidosis in dogs. *Am J Physiol* 253(3 Pt 1): E271–276.
- Gladden LB (2008) A lactic perspective on metabolism. *Med Sci Sports Exerc* 40: 477–85.
- Qvist V, Hagström-Toft E, Moberg E, Sjöberg S, Bolinder J (2007) Lactate release from adipose tissue and skeletal muscle in vivo: defective insulin regulation in insulin-resistant obese women. *Am J Physiol Endocrinol Metab* 292: E709–714.
- Yki-Järvinen H, Bogardus C, Foley JE (1990) Regulation of plasma lactate concentration in resting human subjects. *Metab Clin Exp* 39: 859–64.
- Consoli A, Nurjhan N, Reilly JJ Jr, Bier DM, Gerich JE (1990) Contribution of liver and skeletal muscle to alanine and lactate metabolism in humans. *Am J Physiol* 259(5 Pt 1): E677–684.
- Brecher AS, Lehti MD (1996) A hypothesis linking hypoglycemia, hyperuricemia, lactic acidemia, and reduced gluconeogenesis in alcoholics to inactivation of glucose-6-phosphatase activity by acetaldehyde. *Alcohol* 13: 553–7.
- Stefanovski D, Youn JH, Rees M, Watanabe RM, Ader M, et al. (2012) Estimating hepatic glucokinase activity using a simple model of lactate kinetics. *Diabetes Care* 35: 1015–20.
- Howell RR (1965) The interrelationship of glycogen storage disease and gout. *Arthritis Rheum* 8: 780–5.
- Hosogai N, Fukuhara A, Oshima K, Miyata Y, Tanaka S, et al. (2007) Adipose tissue hypoxia in obesity and its impact on adipocytokine dysregulation. *Diabetes* 56: 901–11.
- Avogaro A, Toffolo G, Miola M, Valerio A, Tiengo A, et al. (1996) Intracellular lactate- and pyruvate-interconversion rates are increased in muscle tissue of non-insulin-dependent diabetic individuals. *J Clin Invest* 98: 108–15.
- Sandqvist MM, Eriksson JW, Jansson PA (2001) Increased lactate release per fat cell in normoglycemic first-degree relatives of individuals with type 2 diabetes. *Diabetes* 50: 2344–8.
- Crandall DL, Fried SK, Francendese AA, Nickel M, DiGirolamo M (1983) Lactate release from isolated rat adipocytes: influence of cell size, glucose concentration, insulin and epinephrine. *Horm Metab Res* 15: 326–9.
- Ye J (2011) Adipose tissue vascularization: its role in chronic inflammation. *Curr Diab Rep* 11: 203–10.
- Björntorp P, Schersten T, Fagerberg SE (1967) Respiration and phosphorylation of mitochondria isolated from the skeletal muscle of diabetic and normal subjects. *Diabetologia* 3: 346–52.
- Kelley DE, He J, Menshikova EV, Ritov VB (2002) Dysfunction of mitochondria in human skeletal muscle in type 2 diabetes. *Diabetes* 51: 2944–50.
- Morino K, Petersen KF, Dufour S, Befroy D, Frattini J, et al. (2005) Reduced mitochondrial density and increased IRS-1 serine phosphorylation in muscle of insulin-resistant offspring of type 2 diabetic parents. *J Clin Invest* 115: 3587–93.
- Kim JY, Hickner RC, Cortright RL, Dohm GL, Houmard JA (2000) Lipid oxidation is reduced in obese human skeletal muscle. *Am J Physiol Endocrinol Metab* 279: E1039–1044.
- Hoeks J, van Herpen NA, Mensink M, Moonen-Kornips E, van Beurden D, et al. (2010) Prolonged fasting identifies skeletal muscle mitochondrial dysfunction as consequence rather than cause of human insulin resistance. *Diabetes* 59: 2117–25.
- Cheng Z, Tseng Y, White MF (2010) Insulin signaling meets mitochondria in metabolism. *Trends Endocrinol Metab* 21: 589–98.
- Holloszy JO (2009) Skeletal muscle “mitochondrial deficiency” does not mediate insulin resistance. *Am J Clin Nutr* 89: 463S–6S.
- Irving BA, Short KR, Nair KS, Stump CS (2011) Nine days of intensive exercise training improves mitochondrial function but not insulin action in adult offspring of mothers with type 2 diabetes. *J Clin Endocrinol Metab* 96: E1137–1141.
- Östergård T, Andersen JL, Nyholm B, Lund S, Nair KS, et al. (2006) Impact of exercise training on insulin sensitivity, physical fitness, and muscle oxidative capacity in first-degree relatives of type 2 diabetic patients. *Am J Physiol Endocrinol Metab* 290: E998–1005.
- Dela F, Stallknecht B (2010) Effect of physical training on insulin secretion and action in skeletal muscle and adipose tissue of first-degree relatives of type 2 diabetic patients. *Am J Physiol Endocrinol Metab* 299: E80–91.
- Pravenec M, Hyakukoku M, Houstek J, Zidek V, Landa V, et al. (2007) Direct linkage of mitochondrial genome variation to risk factors for type 2 diabetes in conplastic strains. *Genome Res* 17: 1319–26.
- Reardon W, Ross RJ, Sweeney MG, Luxon LM, Pembrey ME, et al. (1992) Diabetes mellitus associated with a pathogenic point mutation in mitochondrial DNA. *Lancet* 340: 1376–9.
- van den Ouweland JM, Lemkes HH, Ruitenbeek W, Sandkuijl LA, de Vijlder MF, et al. (1992) Mutation in mitochondrial tRNA(Leu)(UUR) gene in a large pedigree with maternally transmitted type II diabetes mellitus and deafness. *Nat Genet* 1: 368–71.
- Pagel-Langenickel I, Bao J, Pang L, Sack MN (2010) The role of mitochondria in the pathophysiology of skeletal muscle insulin resistance. *Endocr Rev* 31: 25–51.
- Toledo FGS, Menshikova EV, Ritov VB, Azuma K, Radikova Z, et al. (2007) Effects of physical activity and weight loss on skeletal muscle mitochondria and relationship with glucose control in type 2 diabetes. *Diabetes* 56: 2142–7.
- Skov V, Glinborg D, Knudsen S, Tan Q, Jensen T, et al. (2008) Pioglitazone enhances mitochondrial biogenesis and ribosomal protein biosynthesis in skeletal muscle in polycystic ovary syndrome. *PLoS ONE* 3: e2466.
- Watson RT, Kanzaki M, Pessin JE (2004) Regulated membrane trafficking of the insulin-responsive glucose transporter 4 in adipocytes. *Endocr Rev* 25: 177–204.
- Stenbit AE, Tsao TS, Li J, Burcelin R, Geenen DL, et al. (1997) GLUT4 heterozygous knockout mice develop muscle insulin resistance and diabetes. *Nat Med* 3: 1096–1101.
- Ceddia RB, Somwar R, Maida A, Fang X, Bikopoulos G, et al. (2005) Globular adiponectin increases GLUT4 translocation and glucose uptake but reduces glycogen synthesis in rat skeletal muscle cells. *Diabetologia* 48: 132–139.
- Choi CS, Kim YB, Lee FN, Zabolotny JM, Kahn BB et al. (2002) Lactate induces insulin resistance in skeletal muscle by suppressing glycolysis and impairing insulin signaling. *Am J Physiol Endocrinol Metab* 283: E233–240.